

Subclinical ileitis: Diagnostic and performance parameters in a multi-dose mucosal homogenate challenge model

Marie-Anne Paradis, DVM; Connie J. Gebhart, MSc, PhD; Denise Toole, BSc Agr; Gordon Vessie, Diplomate APM; Nathan L. Winkelman, DVM; Sharon A. Bauer, MSc; Jeff B. Wilson, DVM, DVSc, PhD; Carol A. McClure, DVM, MSc, PhD

Summary

A dose titration study was conducted using a *Lawsonia intracellularis* mucosal homogenate model in weaned pigs. Significant negative effects on performance were observed in the absence of positive diagnostic indicators or clinical signs. The results obtained showed that subclinical infection can have a detrimental economic impact on swine herds.

Received: October 27, 2010

Accepted: September 19, 2011

Resumen - Ileitis subclínica: Parámetros de desempeño y diagnóstico en un modelo de prueba de homogeneizado de mucosa en multidosis

Se realizó un estudio de titulación de dosis utilizando un modelo homogeneizado de mucosa de *Lawsonia intracellularis* en cerdos destetados. Se observaron efectos significativamente negativos en el desempeño a pesar de la ausencia de indicadores de diagnóstico positivo ó signos clínicos. Los resultados obtenidos mostraron que la infección subclínica puede tener un impacto económico perjudicial en los hatos porcinos.

Résumé - Iléite sub-clinique: Diagnostic et paramètres de performances dans un modèle d'infection expérimentale utilisant des doses multiples d'un homogénat de muqueuse

Une étude de titration de dose a été menée chez des porcs sevrés à l'aide d'un modèle utilisant *Lawsonia intracellularis* provenant d'un homogénat de muqueuse. Des effets négatifs significatifs sur les performances ont été observés en absence d'indicateurs de diagnostic positif ou de signes cliniques. Les résultats obtenus ont démontré que l'infection sub-clinique peut avoir un impact économique négatif dans les troupeaux porcins.

Porcine proliferative enteropathy (PPE or ileitis) caused by *Lawsonia intracellularis*, an obligate intracellular bacterium, primarily affects the mucosa of the distal ileum.^{1,2} Ileitis has a worldwide distribution, affecting 57% to 100% of herds.^{1,3} Overall, prevalence is underestimated, as some mild or subclinically affected farms go undetected and undiagnosed.⁴ Clinically, ileitis manifests itself in chronic and acute forms.^{1,2} Chronic infection, known as porcine intestinal adenomatosis, occurs in weaned pigs and young growing animals and is characterized by proliferation of mucosal crypt cells in the ileum, jejunum,

and sometimes large intestine. This chronic condition can result in diarrhea, decreased feed intake, and a slower rate of weight gain.^{5,6} Acute infection occurs primarily in mature pigs (> 4 months old) causing proliferative hemorrhagic enteropathy (PHE), which is also characterized by proliferation of the crypt cells. Clinical consequences of PHE include weakness, lethargy, anorexia with bloody diarrhea from intestinal hemorrhage, and sudden death.⁶ In addition to the prominent clinical manifestations of ileitis in swine, a subclinical form of the disease has more recently been recognized and identified both in natural and challenge infection

studies.⁷⁻¹⁰ Subclinical disease caused by *L. intracellularis* infection remains incompletely characterized, but results in enterocyte hyperplasia and reduced performance with few or no clinical signs. The objective of this study was to further define subclinical ileitis by measuring the impact of varying doses of *L. intracellularis* on clinical signs, *L. intracellularis* shedding and seropositivity, performance, and gross and histopathological intestinal changes in weaned pigs.

Materials and methods

Study animals and protocol

The protocol for this study was approved by the Nutreco Agresearch Animal Care and Use Committee.

A total of 144 two-week-old pigs originating from one farrowing unit were weighed and randomly assigned to 24 pens of six pigs each at the beginning of the study (Day -14). Pigs were acclimated for a period of 14 days (Days -14 to 0), challenged with an *L. intracellularis* inoculum on Day 0, and observed for a period of 21 to 22 days, at which time each pig was euthanized and necropsied. The pigs to be euthanized (three pigs per pen) on Day 21 or 22 were randomly pre-assigned on Day -14.

Ante mortem diagnostic testing

Blood and fecal samples were collected from two randomly selected pigs in each pen

MAP: Elanco Animal Health, St-Jean-sur-Richelieu, Québec, Canada.

CJG: University of Minnesota, St Paul, Minnesota.

DT: Nutreco Agresearch, Burford, Ontario, Canada.

GV: Elanco Animal Health, Guelph, Ontario, Canada.

NLW: Swine Services Unlimited Inc, Rice, Minnesota.

SAB, JBW: Ontario Veterinary College, University of Guelph, Guelph, Ontario, Canada.

CAM: Atlantic Veterinary College, University of Prince Edward Island, Charlottetown, Prince Edward Island, Canada.

Corresponding author: Dr Marie-Anne Paradis, 7-624 boul du Séminaire Nord, St-Jean-sur Richelieu, Québec, J3B 7B4 Canada; Tel: 450-349-9539; Fax: 450-349-9541; E-mail: paradis_marie_anne@elanco.com.

Dr Wilson has served as a paid consultant to Elanco Animal Health.

This study was funded by Elanco, Division Eli Lilly Canada Inc.

This article is available online at <http://www.aasv.org/shap.html>.

Paradis MA, Gebhart CJ, Toole D, et al. Subclinical ileitis: Diagnostic and performance parameters in a multi-dose mucosal homogenate challenge model. *J Swine Health Prod.* 2012;20(3):137-141.

on Day -14, Day 14, and at the end of the study (Day 21 or 22). Serum samples were analyzed for antibody to *L intracellularis* by serum immunoperoxidase monolayer assay (IPMA),¹¹ and fecal swabs were tested for *L intracellularis* by polymerase chain reaction (PCR) using previously described methods.^{11,12}

Study design and inoculum preparation

The experimental design was a randomized complete block with blocking on body weight. There were six treatment groups with four pens of six pigs per treatment. Each study group was treated on Day 0 with a different dose of *L intracellularis* inoculum: Treatment A (nonchallenged, control) and Treatments B, C, D, E, and F which were challenged with serial 10-fold dilutions resulting in doses of approximately 10^4 (F) to 10^8 (B) organisms per pig. Preparation of inoculum was performed similarly to Winkelman et al.¹³ and samples were cultured to confirm absence of other enteric pathogens, including *Brachyspira* species, *Salmonella* serovar Choleraesuis, and β -hemolytic *Escherichia coli*. Control pigs were treated first with pure sucrose phosphate glutamate (SPG), and the five challenged groups were treated beginning with the most dilute dose and progressing up to higher doses.

Housing, feeding, clinical scoring, and performance measurements

Each pen contained a feeder and a nipple drinker and had completely slatted floors and solid PVC partitions. Feed and water were provided ad libitum. A commercial, nonmedicated diet was fed during the entire study. Clinical scores for attitude, abdominal appearance, fecal consistency, and fecal blood were recorded for each pig three times per week, from Day 7 to Day 21.

All pigs were individually weighed on Days -14, 0, 7, 14, and 21 or 22. Feed intake was calculated on each weigh day.

Postmortem diagnostics

Postmortem examination was conducted by a pathologist who was blinded to treatments for every pig that died or was euthanized during the study. In each pig, the jejunum, ileum, cecum, and colon were opened and scored for consistency of content and presence of blood, mucosal thickness and necrosis, gross diagnosis of ileitis, and length of lesions. All sections were stained with hematoxylin and eosin (H&E). Sections of ileum were stained by the Warthin-Starry (WS)

technique to identify intracellular organisms and by immunohistochemistry (IHC) for *L intracellularis* using an immunoperoxidase staining technique incorporating a specific polyclonal antibody. Sections were scored for mucosal epithelial proliferation, presence of intraepithelial bacteria, proprial inflammation, mucosal necrosis, and crypt abscessation.

Statistical analysis

All analyses were conducted using analysis of variance (ANOVA) with the pen as the experimental unit and using a statistical significance level of $P < .05$, unless otherwise noted. Tukey's pairwise comparisons were used for significant differences between treatment groups.¹⁴ Categorical data were transformed into arcsine of the square root of proportions per pen and then analyzed using ANOVA.

Results

Ante mortem diagnostic testing and inoculum

All tested animals were negative for *L intracellularis* both by fecal PCR and serological testing on Day -14 (Table 1). By Day 14, approximately 15% of pigs in Treatments B and C were PCR-positive, while the other treatment groups remained PCR-negative. On Days 21 and 22, fecal samples from 12.5% to 83.6% of the challenged groups were PCR-positive, and 12.5% to 98.0% of challenged pigs were seropositive for *L intracellularis* antibodies. More pigs in Treatments C and D were PCR-positive for *L intracellularis* than in Treatment A (nonchallenged controls) ($P < .01$), and more pigs in Treatments B, C, and D were seropositive than in either Treatment E or the nonchallenged controls ($P < .01$). No animals in the nonchallenged control group became positive either by fecal PCR or by IPMA. No contaminating viruses or parasites were identified in the inoculum. Bacterial culture was negative for *Brachyspira* species, *Salmonella*, and β -hemolytic *E coli* for all inoculum dilutions.

Clinical scores

Clinical scores for abdominal appearance, pig attitude, and fecal blood were predominantly normal for all treatments across all observation periods ($P > .05$) (data not shown). There was no significant difference in mean fecal consistency score between treatment groups until Day 14 (Table 2). Between sampling days 14 and 18, there was a numerical dose response between the number of organisms administered and the mean

fecal consistency score for the pens. Fecal consistency scores were significantly higher than those of the nonchallenged controls in Treatments B and C on Day 14 ($P < .01$), in all challenged groups on Day 16 ($P = .01$), and in Treatments B, C, and D on Day 18 ($P < .01$) (Table 2).

Performance

Performance results are presented in Tables 3 and 4. Prior to challenge, growth performance did not differ between treatments (data not shown). Growth rate was numerically lower in challenged groups than in controls Days 0 to 6 (Table 4). Average daily gain was significantly lower in Treatment B than in the nonchallenged controls for Days 7 to 13 ($P < .05$). On Day 14, pigs in Treatments B and E weighed significantly less (by approximately 2 kg) than did the nonchallenged controls (Table 3). Average daily gain was significantly lower than controls ($P < .01$) in all challenged groups for Days 14 to 21 and 22 and Days 0 to 21 and 22 (Table 4), paralleling the lower body weights observed during these periods. Average daily feed intake was also significantly lower in all challenged groups than in the nonchallenged controls for Days 7 to 13, Days 14 to 21 and 22, and Days 0 to 21 and 22 ($P < .01$). There was a significantly poorer ($P < .01$) feed conversion ratio over the entire post-challenge period (Days 0 to 21 and 22) with increasing inoculum dose. Generally, performance was poorer (Days 0 to 21 and 22) for all performance measures with increasing inoculum doses.

Mortality

Mortality was distributed across all treatment groups, with no significant difference between treatments ($P > .05$) (data not shown). Thirteen of the 17 pigs that died during the study were diagnosed with K88 *E coli* septicemia during the early postchallenge period (Days 2 to 5); affected pigs were reported in each treatment group, including the nonchallenged control group. Two pigs (one from Treatment F on Day 14 and one from Treatment C on Day 19) died of nonspecific intestinal disease. On Days 20 and 22, respectively, one pig in Treatment B and one in Treatment C died of ileitis.

Postmortem diagnostics

Successful disease challenge was confirmed by postmortem diagnostic testing (Table 5). Between 8% (Treatment F) and 33% (Treatment D) of the pigs in challenged pens had gross lesions consistent with *L intracellularis* infection at necropsy on Days 21 and 22, and no lesions were observed in the nonchal-

Table 1: Mean proportions of animals positive for *Lawsonia intracellularis* (LI) by fecal PCR and serum IPMA testing in nursery pigs challenged on Day 0* with a mucosal homogenate containing LI

Treatment	No. of LI organisms per pig	Fecal PCR			Serum IPMA		
		Day -14	Day 14	Days 21-22	Day -14	Day 14	Days 21-22
A	0	0.000	0.000	0.000 ^a	0.000	0.000	0.000 ^a
B	2.4 × 10 ⁸	0.000	0.151	0.500 ^{ab}	0.000	0.125	0.875 ^{bc}
C	7.2 × 10 ⁷	0.000	0.142	0.836 ^b	0.000	0.000	0.980 ^c
D	2.2 × 10 ⁶	0.000	0.000	0.750 ^b	0.000	0.000	0.750 ^{bc}
E	3.8 × 10 ⁵	0.000	0.000	0.125 ^a	0.000	0.125	0.125 ^a
F	3.2 × 10 ⁴	0.000	0.000	0.500 ^{ab}	0.000	0.125	0.375 ^{ab}
SEM	NA	NA	0.073	0.144	NA	0.099	0.125
P	NA	NA	.49	< .01	NA	.72	< .01

* Each treatment group included four pens of six pigs (total 24 pigs). Pigs were weaned at 14 days of age (Day -14), challenged with varying doses of LI at 28 days of age (Day 0), and euthanized on Day 21 or 22 (half of each group on each day). Fecal and blood samples were collected from two pigs per pen for fecal PCR and serological testing for LI (total eight pigs per treatment). Sera were tested for antibodies to LI by IPMA. An effort was made to collect samples from the same two pigs per pen on each occasion. The unit of analysis was the pen.

^{abc} Values in a column with no common superscript are significantly different ($P < .01$; ANOVA).

PCR = polymerase chain reaction; IPMA = immunoperoxidase monolayer assay; NA = not applicable.

Table 2: Mean fecal consistency scores in groups of pigs weaned at 14 days of age and inoculated 14 days later (Day 0) with varying doses of a mucosal homogenate containing *Lawsonia intracellularis**

Treatment	Day 7	Day 9	Day 11	Day 14	Day 16	Day 18	Days 21-22
A	0.10	0.20	0.08	0.08 ^a	0.00 ^a	0.00 ^a	0.17
B	0.53	0.57	0.70	1.34 ^b	1.02 ^b	0.94 ^b	0.68
C	0.70	0.64	0.56	0.93 ^b	0.89 ^b	0.83 ^b	0.81
D	0.42	0.52	0.45	0.37 ^a	0.33 ^b	0.65 ^b	0.83
E	0.78	0.59	0.88	0.43 ^a	0.42 ^b	0.29 ^a	0.33
F	0.29	0.43	0.38	0.18 ^a	0.28 ^b	0.29 ^a	0.85
SEM	0.22	0.27	0.22	0.20	0.19	0.12	0.19
P	.32	.87	.24	< .01	.01	< .01	.10

* Each treatment group included four pens of six pigs (total 24 pigs). Score of 0, normal feces; 1, moderate diarrhea; 2, liquid feces. Inoculum doses for Treatments B through F shown in Table 1. Treatment A pigs were nonchallenged controls. Pen was the unit of analysis.

^{ab} Values in a column with no common superscript are significantly different ($P < .01$; ANOVA).

lenged controls. More positive animals were identified by H&E scores (31% to 61%), WS scores (33% to 54%), and IHC scores (62% to 78%) than by gross pathology (8% to 33%). The most sensitive indicator was IHC, with 62% to 78% of the challenged pigs having scores that were significantly different from those of the controls ($P < .01$). The proportions of pigs in which the ileum was histologically positive for ileitis were significantly greater ($P < .05$) in Treatments C and D than in the nonchallenged control group. When all four post mortem diagnostic measures of ileitis were considered, the proportion of pigs positive was highest in Treatment D ($P < .05$) when compared to the nonchallenged controls.

Discussion

The present study demonstrates both induction of subclinical ileitis after inoculation with a mucosal homogenate containing *L. intracellularis* and a dose response to the number of organisms in the inoculum, creating a spectrum of illness from subclinical disease at the lowest inoculum dosage to clinical disease at higher doses and even death of some infected pigs. By the end of the study, impaired performance was observed at all doses of *Lawsonia* organisms (Treatments B to F). The consistency of poor performance in the challenged groups made performance parameters the most sensitive indicators to identify the disease process experienced by these pigs. The impact of the subclinical form of ileitis on growth was the most remarkable observation of this study. Even at the lowest inoculum dose (Treatment F), average daily gain during the trial period was 37% lower than that of the nonchallenged pigs, and feed conversion was 27% higher. There was no clinical difference in mean fecal consistency scores between Treatments E and F (the two lowest doses of inoculum administered) and the nonchallenged control groups except for one day (Day 16). In a field situation, such an observation could easily go undetected. This is consistent with an observational study⁸ in which it was concluded that infection with *L. intracellularis* can result in subclinical disease, demonstrated by poor performance of pigs that did not have diarrhea. Fecal shedding of *L. intracellularis* was detected in some pigs, many of these subclinically infected. Although no clinical signs of disease were observed, histopathological evidence of *L. intracellularis* infection was found. This may be extremely important

Table 3: Mean body weight in groups of pigs weaned at 14 days of age and inoculated 14 days later (Day 0) with varying doses of a mucosal homogenate containing *Lawsonia intracellularis**

Treatment	Body weight (kg)			
	Day 0	Day 7	Day 14	Day 21-22
A	7.68	9.19	11.80 ^a	16.51 ^a
B	7.29	8.23	9.47 ^b	10.93 ^b
C	7.28	8.59	10.35 ^{ab}	11.95 ^b
D	7.35	8.91	10.83 ^{ab}	12.84 ^b
E	7.23	8.14	9.93 ^b	12.34 ^b
F	7.17	8.41	10.31 ^{ab}	13.00 ^b
SEM	0.11	0.24	0.33	0.53
P	.07	.05	< .01	< .01

* Each treatment group included four pens of six pigs (total 24 pigs) on Days -14 and 0. During Days 2 to 5, 13 pigs died of K88 *Escherichia coli* septicemia, evenly distributed among the treatment groups; one treatment C pig and one treatment F pig died of nonspecific gastrointestinal disease on Days 14 and 19, respectively; and one treatment B pig and one treatment C pig died of ileitis on Days 20 and 22, respectively. Inoculum doses for Treatments B through F are shown in Table 1. Treatment A pigs were nonchallenged controls. The unit of analysis was the pen.

^{ab} Mean values in a column with no common superscript are significantly different ($P < .01$; ANOVA).

for the economics of swine farming. Not only do subclinically infected pigs perform suboptimally, but their feces may infect other pigs in the herd.¹⁵

Histological evidence of *L. intracellularis* infection concurrent with minimal clinical effects suggests that the inoculum resulted in subclinical infection for many pigs in this study. Severity of ileal lesions, observed grossly and histologically, increased with dose of *L. intracellularis*. However, peak gross pathology scores (consistency of contents, mucosal thickness, mucosal necrosis, and length of lesions) and peak histopathology scores (H&E, WS, and IHC) occurred unexpectedly in Treatments C and D rather than in Treatment B, which received the highest dose of organisms. This could be explained by the fact that in Treatment B, there was a shorter incubation period and course of disease so that lesions were resolving and appeared less severe at necropsy. Another study¹⁶ noted that detecting resolving lesions was difficult, as the *L. intracellularis* were no longer found in the enterocytes.

Most deaths in this study were caused by *E. coli* K88 infection in the first 5 days post

Table 4: Mean measures of performance in groups of pigs weaned at 14 days of age and inoculated 14 days later (Day 0) with varying doses of a mucosal homogenate containing *Lawsonia intracellularis**

Treatment	Days 0-6			Days 7-13			Days 14-21 (22)			Days 0-21 (22)		
	ADG (kg)	ADI (kg)	F:G (kg:kg)	ADG (kg)	ADI (kg)	F:G (kg:kg)	ADG (kg)	ADI (kg)	F:G (kg:kg)	ADG (kg)	ADI (kg)	F:G (kg:kg)
A	0.185	0.385	2.12	0.373 ^a	0.606 ^c	1.64	0.629 ^c	0.948 ^c	1.51	0.396 ^c	0.647 ^c	1.63 ^c
B	0.110	0.339	3.50	0.176 ^b	0.439 ^d	2.83	0.180 ^d	0.543 ^d	4.44	0.155 ^d	0.441 ^d	2.92 ^d
C	0.159	0.368	2.47	0.252 ^{ab}	0.469 ^d	1.87	0.163 ^d	0.542 ^d	13.71	0.190 ^d	0.460 ^d	2.51 ^{de}
D	0.173	0.374	2.75	0.275 ^{ab}	0.491 ^d	1.83	0.267 ^d	0.654 ^d	3.53	0.237 ^d	0.507 ^d	2.24 ^{de}
E	0.120	0.331	2.92	0.256 ^{ab}	0.442 ^d	1.98	0.322 ^d	0.662 ^d	2.09	0.234 ^d	0.482 ^d	2.10 ^e
F	0.153	0.357	2.56	0.238 ^{ab}	0.453 ^d	1.93	0.363 ^d	0.730 ^d	2.06	0.249 ^d	0.511 ^d	2.07 ^e
SEM	0.026	0.016	0.34	0.035	0.027	0.32	0.050	0.052	4.62	0.025	0.024	0.18
P	.32	.18	.14	.03	< .01	.19	< .01	< .01	0.45	< .01	< .01	< .01

* Each treatment group included four pens of six pigs (total 24 pigs) on Day -14 and 0. Inoculum doses for Treatments B through F shown in Table 1. Treatment A pigs were nonchallenged controls. Half of the pigs in each group were euthanized on Day 21 and the remaining pigs on Day 22. The unit of analysis was the pen.

^{ab} Mean values in a column with no common superscript are significantly different ($P < .05$; ANOVA).

^{cde} Mean values in a column with no common superscript are significantly different ($P < .01$; ANOVA).

ADG = average daily gain; ADI = average daily intake; F:G = feed-to-gain ratio.

Table 5: Mean proportions of pigs exhibiting diagnostic measures of ileitis 21 or 22 days post challenge with a mucosal homogenate containing *Lawsonia intracellularis* (LI)*

Treatment	No. of LI organisms per pig†	Gross diagnosis	H&E‡	WS§	IHC¶
		Score > 0	Score > 0	Score > 0	Score > 0
A	0	0.00 ^a	0.00 ^a	0.00 ^c	0.00 ^c
B	2.4 × 10 ⁸	0.25 ^{ab}	0.33 ^{ab}	0.33 ^{cd}	0.62 ^d
C	7.2 × 10 ⁷	0.25 ^{ab}	0.57 ^b	0.46 ^d	0.66 ^d
D	2.2 × 10 ⁶	0.33 ^b	0.61 ^b	0.54 ^d	0.78 ^d
E	3.8 × 10 ⁵	0.13 ^{ab}	0.31 ^{ab}	0.38 ^d	0.63 ^d
F	3.2 × 10 ⁴	0.08 ^{ab}	0.34 ^{ab}	0.42 ^d	0.67 ^d
SEM	NA	0.07	0.12	0.08	0.07
P	NA	.03	.03	< .01	< .01

* Each treatment group included four pens of six pigs. Pigs were weaned at 14 days of age and inoculated with LI 14 days later (Day 0). Treatment A pigs were nonchallenged controls. Samples were collected at euthanasia 21 or 22 days post challenge (half of each group euthanized each day). Gross diagnosis: 0 = absence of gross lesions of PPE, 1 = presence of gross lesions of PPE; H&E: 0 = no lesions, 1 = mild, 2 = moderate, 3 = marked; WS: 0 = none, 1 = few definitive intraepithelial bacteria in some crypts, 2 = abundant definitive intraepithelial bacteria in some crypts, 3 = abundant definitive intraepithelial bacteria in many crypts; IHC: 0 = no specific staining, 1 = staining in < 25% of crypts, 2 = staining in 25% to < 50% of crypts, 3 = staining in 50% to < 75% of crypts, 4 = staining in ≥ 75% of crypts.

† 40 mL of sucrose-phosphate-glutamate buffer alone was administered by stomach tube to each Treatment A pig, and 40 mL of inoculum was similarly administered to each challenged pig.

‡ Proportion of pigs showing histological evidence of porcine proliferative enteropathy in small intestinal tissue (combined jejunal and ileal segments) stained with H&E.

§ Proportion of pigs in which organisms characteristic of LI were observed in ileal sections stained with WS stain.

¶ Proportion of pigs in which ileal sections were positive for LI by specific IHC staining.

^{ab} Mean values within a column with no common superscript are significantly different ($P < .05$; ANOVA).

^{cd} Mean values within a column with no common superscript are significantly different ($P < .01$; ANOVA).

H&E = hemotoxylin and eosin; WS = Warthin-Starry stain; IHC = immunohistochemistry; PPE = porcine proliferative enteropathy.

inoculation. As *E. coli* K88 was not cultured from the inocula and the nonchallenged control group was also affected, it is likely that the source of this pathogen was the pigs themselves. Little information was contributed to the study by these pigs, which died before most of the study measurements were made. The end result is that there may have been a decrease in statistical power, but that does not change the effects observed.

Although subclinical ileitis was induced in previous studies, the range of inoculum doses, the number of pigs included, and the measurements were more extensive in the current study. The results confirm that

subclinical infection can have a significant impact on growth performance, and that subclinically infected pigs can serve as a source of infection for other animals in the herd. Moreover, a significantly smaller proportion of these subclinically affected animals are positive by routine diagnostic testing methods such as seroconversion and fecal PCR.

Implications

- Subclinical infection caused by *L. intracellularis* can have a significant impact on growth performance.
- Subclinically infected pigs can serve as a

source of infection for other pigs in the herd.

- Routine diagnostic methods such as seroconversion and fecal PCR may be negative in subclinically infected pigs.

References

1. Lawson GHK, Gebhart CJ. Proliferative enteropathy. *J Comp Pathol.* 2000;122:77–100.
2. McOrist S, Gebhart CJ. Proliferative enteropathies. In: Straw BE, Zimmerman JJ, D’Allaire S, Taylor DJ, eds. *Diseases of Swine*. 9th ed. Ames, Iowa: Iowa State University Press; 2006:727–737.
3. McOrist S, Barcellos DE, Wilson RJ. Global patterns of porcine proliferative enteropathy. *Pig J.* 2003;51:26–35.
4. Smith SH, McOrist S, Green LE. Questionnaire survey of proliferative enteropathy on British pig farms. *Vet Rec.* 1998;142:690–693.
5. Roberts L, Lawson GHK, Rowland AC, Laing AH. Porcine intestinal adenomatosis and its detection in a closed pig herd. *Vet Rec.* 1979;104:366–368.
6. Winkelman NL. Ileitis: an update. *Comp Cont Ed Pract Vet.* 1996;18:S19–S25.
7. Guedes RMC, Winkelman NL, Gebhart CJ. Relationship between the severity of porcine proliferative enteropathy and the infectious dose of *Lawsonia intracellularis*. *Vet Rec.* 2003;153:432–433.
8. Jacobson M, Hård af Segerstad C, Gunnarsson A, Fellström C, de Verdier Klingenberg K, Wallgren P, Jensen-Waern M. Diarrhoea in the growing pig – a comparison of clinical, morphological and microbial findings between animals from good and poor performance herds. *Res Vet Sci.* 2003;74:163–169.
9. Barker IK, Van Dreumel AA, Palmer N. The alimentary system. In: Jubb KVF, Kennedy PC, Palmer N, eds. *Pathology of Domestic Animals*. 4th ed. San Diego, California: Academic Press; 1993:229–233.
10. Gogolewski RP, Cook RW, Batterham ES. Sub-optimal growth associated with porcine intestinal adenomatosis in pigs in nutritional studies. *Aust Vet J.* 1991;68:406–408.
11. Guedes RM, Gebhart CJ, Deen J, Winkelman NL. Validation of an immunoperoxidase monolayer assay as a serologic test for porcine proliferative enteropathy. *J Vet Diagn Invest.* 2002;14:528–530.
12. Guedes RM, Gebhart CJ, Winkelman NL, Mackie-Nuss RA, Marsteller TA, Deen J. Comparison of different methods for diagnosis of porcine proliferative enteropathy. *Can J Vet Res.* 2002;66:99–107.
13. Winkelman NL, Crane JP, Elfring GD, Kratzer DD, Meeuwse DM, Dame KJ, Buckham SL, Gebhart CJ. Lincomycin-medicated feed for the control of porcine proliferative enteropathy (ileitis) in swine. *J Swine Health Prod.* 2002;10:107–111.
14. Neter J, Kutner MH, Nachtsheim CJ, Wasserman W. *Applied Linear Statistical Models*. 4th ed. Boston, Massachusetts: WCB McGraw-Hill; 2004.
15. van der Heijden HMJF, Bakker J, Elbers ARW, Vos JH, Weyns A, de Smet M, McOrist S. Prevalence of exposure and infection of *Lawsonia intracellularis* among slaughter-age pigs. *Res Vet Sci.* 2004;77:197–202.
16. Ladinig A, Sommerfeld-Stur I, Weissenböck H. Comparative evaluation of diagnostic methods for *Lawsonia intracellularis* infection in pigs, with emphasis on cases lacking characteristic lesions. *J Comp Pathol.* 2009;140:140–148.

